Highly sensitive screening test for the detection of equine anti-Anaplasma antibodies
Based on recombinant antigen
Fully automatable

Technical data

Antigen: Recombinantly produced and purified Anaplasma phagocytophilum antigen
Calibration: Semiquantitative evaluation using ratio values:
  Extinction value of the sample over the extinction value of the calibrator
Result interpretation: EUROIMMUN recommends interpreting results as follows:
  Ratio < 0.8: negative
  Ratio ≥ 0.8 to < 1.1: borderline
  Ratio ≥ 1.1: positive
Sample dilution: Equine serum or plasma, 1:101 in sample buffer
Reagents: Ready for use, with the exception of the wash buffer (10x), colour-coded solutions
Test procedure: 30 min (37°C) / 30 min (37°C) / 15 min (room temperature), fully automatable
Measurement: 450 nm, reference wavelength between 620 and 650 nm
Test kit format: 96 break-off wells, kit includes all necessary reagents
Order no.: EI 220m-9601 GE

Clinical significance

Anaplasmosis is a disease which is transmitted to animals and humans by ticks of the Ixodes genus. It is caused by Anaplasma (A.) phagocytophilum (formerly: Ehrlichia phagocytophila, Ehrlichia equi). The disease was first described in a horse in the USA in 1969. A. phagocytophilum is distributed worldwide, its prevalence depends on the occurrence of the transmitting vectors. The seroprevalence is given as 17 to 43%, but amounts to up to 50% in endemic areas in healthy horses. There are different names for the clinical image of an infection with A. phagocytophilum in horses: granulocytic ehrlichiosis (obsolete), equine granulocytic anaplasmosis, and, simply, and most frequently used: anaplasmosis. A. phagocytophilum is a gram-negative, obligate intracellular bacterium which attacks mostly neutrophilic granulocytes, but also, in rare cases, eosinophilic granulocytes.

The clinical symptoms of equine anaplasmosis are reduced general condition with fever, anorexia, lethargy, oedema of the limbs, petechia, icterus, reluctance to physical activity and ataxia. Older horses show clearer symptoms than younger ones. Borreliosis should be excluded by differential diagnosis. There are no vaccines available.

Antibodies against A. phagocytophilum occur in the serum of infected animals after 7 to 14 days. Different techniques, such as ELISA or indirect immunofluorescence (IIF), are used for the serological detection of antibodies. It should be noted that many horses that show specific antibodies against A. phagocytophilum are not clinically conspicuous. For diagnosis, it is hence necessary to investigate two consecutive blood samples. A twofold titer increase or a seroconversion are diagnostically relevant. If the first blood sample tests negative, a second sample should be examined after two weeks in cases of suspected anaplasmosis since horses do not produce antibodies in the early phase of infection.
Application

The direct detection of *A. phagocytophilum* by staining or culture is possible, but not conducted routinely since these methods are too complicated for screening diagnostics. PCR is commonly used. The sensitivity of the direct detection depends on the phase of infection as there are “silent phases” when the anaplasms are virtually not detectable in the blood. It is only in reproductive phases (fever attacks) that a reliable direct detection is possible. It is compulsory to use whole blood for PCR. Therefore, serological detection of antibodies is the method of choice when it comes to laboratory diagnosis of equine granulocytic anaplasmosis. Owing to the use of a specific recombinant antigen, the Anti-Anaplasma phagocytophilum ELISA Horse (IgG) has a high specificity and very high sensitivity.

Principle of the test

The ELISA test kit provides a semiquantitative in vitro assay for equine antibodies of the IgG class against *Anaplasma phagocytophilum* antigens in serum or plasma. The test kit contains microtiter strips each with 8 break-off reagent wells coated with recombinantly produced and purified *Anaplasma phagocytophilum* antigen. In the first reaction step, diluted samples are incubated in the wells. In the case of positive samples, specific IgG antibodies (also IgA and IgM) will bind to the antigens. To detect the bound antibodies, a second incubation is carried out using an enzyme-labelled anti-horse IgG (enzyme conjugate) catalysing a colour reaction.

Reproducibility

The reproducibility was investigated by determining the intra- and inter-assay coefficients of variation using three sera. The intra-assay CVs are based on 20 determinations and the inter-assay CVs on 4 determinations performed in 6 different test runs.

<table>
<thead>
<tr>
<th>Serum</th>
<th>Mean value (Ratio)</th>
<th>CV (%)</th>
<th>Mean value (Ratio)</th>
<th>CV (%)</th>
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<td>3.6</td>
<td>2.2</td>
<td>3.6</td>
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<table>
<thead>
<tr>
<th>Precharacterisation (IIFT)</th>
<th>n=59</th>
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<tbody>
<tr>
<td>positive</td>
<td>23</td>
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<tr>
<td>borderline</td>
<td>3</td>
</tr>
<tr>
<td>negative</td>
<td>3</td>
</tr>
</tbody>
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Sensitivity and specificity

59 randomly selected horse sera were investigated using the Anti-Anaplasma phagocytophilum ELISA Horse (IgG) and a commercial immunofluorescence test. The test results were compared and showed a sensitivity of 100 % and a specificity of 90 % (borderline sera were not included in the calculation).

Literature